

Fig. 1 (color online) 0 Gy (a), 1 Gy (b), 4 Gy (c). Photomicrographs of sections stained with H&E (magnification 200 ×): sg, spermatogonia; sc, spermatocytes; esd, early spermatids; lsd, late spermatids; lc, Leydig cell. Severe degeneration of spermatogenic cells in the seminiferous tubule (\*), arrows indicate Sertoli cells in the seminiferous tubule (c). Representative photomicrographs of TUNEL staining: 0 Gy (e), 1 Gy (f), 4 Gy (g). Arrows indicate TUNEL-positive cells in the seminiferous tubule (magnification 400 ×): sg, spermatogonia; sc, spermatocytes; esd, early spermatids; lsd, late spermatids. Testicular damage as evaluated by the Johnsen score (d). Histogram of apoptotic-positive spermatogenic cells from mouse testis (h). Values represent the average  $\pm$  S.E.M. Asterisks indicate a statistically significant difference from control: \*\*\* $P < 0.001$  on one-way ANOVA with Duncan's post hoc analysis.

# Reference

[1] H. Zhang, B. Liu, Q. Zhou, et al., *Int. J. Androl*, 29(2006)592.

## 3 - 63 Toxicity of Mitochondrial Singlet Oxygen Inducer on Zebrafish Embryo\*

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Mitochondrial singlet oxygen is a potential signal molecule to regulate mitochondrial biogenesis<sup>[1]</sup>. However, it is hard to distinguish mitochondrial ROS from cytoplasmic ROS in vitro. To overcome this issue, we recently developed a specific mitochondrial singlet oxygen inducer-Mitochondros. The toxicity of Mitochondros has been verified in vitro. However, the in vivo toxicity of mitochondros has not been investigated. Here we employed the zebrafish embryo to study the toxicity of mitochondros in vivo.

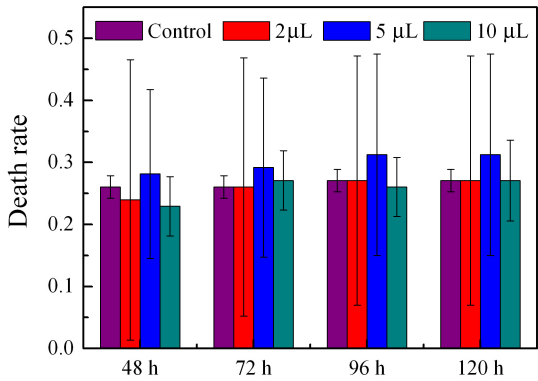


Fig. 1 (color online) The death rate of zebrafish embryo within 120 hours after exposure to various concentration of Mitochondros.

# Reference

[1] R. J. Mailloux, *Redox Biol*, 4(2015)381.

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